

# Stabilization of the retinal vascular network by reciprocal feedback between blood vessels and astrocytes

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## Summary

Development of the retinal vasculature is controlled by a hierarchy of interactions among retinal neurons, astrocytes and blood vessels. Retinal neurons release platelet-derived growth factor (PDGFA) to stimulate proliferation of astrocytes, which in turn stimulate blood vessel growth by secreting vascular endothelial cell growth factor (VEGF). Presumably, there must be counteractive mechanisms for limiting astrocyte proliferation and VEGF production to prevent runaway angiogenesis. Here, we present evidence that the developing vessels provide feedback signals that trigger astrocyte differentiation – marked by cessation of cell division, upregulation of glial fibrillary acidic protein (GFAP) and downregulation of VEGF. We prevented

retinal vessel development by raising newborn mice in a high-oxygen atmosphere, which leads, paradoxically, to retinal hypoxia (confirmed by using the oxygen-sensing reagent EF5). The forced absence of vessels caused prolonged astrocyte proliferation and inhibited astrocyte differentiation *in vivo*. We could reproduce these effects by culturing retinal astrocytes in a low oxygen atmosphere, raising the possibility that blood-borne oxygen itself might induce astrocyte differentiation and indirectly prevent further elaboration of the vascular network.

Keywords: Astrocyte, Retina, Blood vessel, PDGF-A, Oxygen, Transgenic mice

## Introduction

Retinal astrocyte development is believed to be a tightly controlled process. Astrocyte development in the retina is similar to that in the mammalian brain. Astrocytes are derived from the neural plate and differentiate into astrocytes (Srinivasan & D'Amico, 1987; Watanabe & Raff, 1988; Li & Green, 1989; Saucedo et al., 1999). The retinal astrocyte population is regulated by platelet-derived growth factor (PDGF) and vascular endothelial cell growth factor (VEGF), which are secreted by retinal neurons and endothelial cells, respectively (Jiang et al., 1995; Zhang & Srinivasan, 1997; Fruttiger, 2002). The reciprocal mechanism by which retinal astrocytes induce endothelial cell growth is mediated by the astrocyte-derived basic fibroblast growth factor (bFGF), which is secreted by astrocytes and binds to the PDGF receptor, leading to increased proliferation of endothelial cells (D'Amico et al., 2002). Retinal astrocytes also secrete vascular endothelial cell growth factor (VEGF), which stimulates endothelial cell proliferation and migration (Albright et al., 1995; Srinivasan et al., 1995; Pierce et al., 1996; Pridmore et al., 1997). In the adult retina, the endothelial cell population is maintained by the local production of VEGF (D'Amico et al., 2002; Gehring et al., 2003). The development of the retinal vascular network is regulated by a hierarchy of interactions among retinal neurons, astrocytes and blood vessels. Retinal neurons release PDGF to stimulate proliferation of astrocytes, which in turn stimulate blood vessel growth by secreting VEGF. Presumably, there must be counteractive mechanisms for limiting astrocyte proliferation and VEGF production to prevent runaway angiogenesis. Here, we present evidence that the developing vessels provide feedback signals that trigger astrocyte differentiation – marked by cessation of cell division, upregulation of glial fibrillary acidic protein (GFAP) and downregulation of VEGF. We prevented

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ence of gliosis, decrease in the abundance of GFAP in the  
A likely explanation is that the hypoxic environment of the  
dendritic bodies of astrocytes is a result of the hypoxic  
environment of the central nervous system (Clarke and  
Farrington, 2003). Nevertheless, the hypoxic environment is  
not the only factor that regulates astrocyte differentiation.

#### Retinal oxygen tension measured with the hypoxia marker EF5

In order to determine the oxygen tension in the retina, we  
used the EF5 hypoxia marker (Koch, 2002). Mice

om ic  $\alpha$  l i e , b i e m a i e d e l e a e d h r i g h i h e  
 $\alpha$  l i e e i d (Fig. 5I).

## Discussion

A l f e e a c h h a f  $\alpha$  e d l c a l f a c h a c l b l d  
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(Malenka and Simons, 1998). The effect, however, may be due to the fact that different cell types are more likely to be found in different cell types.

How does the frequency of action potentials affect the CNS? Action potentials are generated by the brain and are carried by all GFAP-positive cells, which are GFAP-positive (Bigami and Dahl, 1974). It is possible that action potentials are carried by different cell types, such as the cell bodies, and also by the cell bodies, because the frequency of high-frequency action potentials is affected by the frequency of action potentials. Therefore, the frequency of action potentials is affected by the frequency of action potentials.

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